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(21) International Application Number: PCT/US93/10522 (22) International Filing Date: 3 November 1993 (03.11.93) (30) Priority data: 973,307 9 November 1992 (09.11.92) US (71) Applicant: THE UNITED STATES GOVERNMENT as represented by THE SECRETARY OF THE DEPARTMENT OF HEALTH AND HUMAN SERVICES (US/US); National Institute of Health, Building 31, Room 5A50, 9000 Rockville Pike, Bethesda, MD 20878 (US). (72) Inventors: ANDERSON, W., French ; 960 Winston Avenue, San Marino, CA 91108 (US). BALTRUCKI, Leon, F. ; 14002 Cove Lane, Apt. 203, Rockville, MD 20851 (US). MASON, James, M. ; 14008 Chestnut Court, Laurel, MD 20707 (US).	(74) Agents: OLSTEIN, Elliot, M. et al.; Carella, Byrne, Gilfillan, Cecchi, Stewart & Olstein, 6 Becker Road, Roseland, NJ 07068 (US). (81) Designated States: AU, CA, JP, European patent (A), CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, PT, SE). Published <i>With international search report.</i>	
(54) Title: TARGETABLE VECTOR PARTICLES (57) Abstract A vector particle (e.g., a retroviral vector particle) containing a chimeric envelope includes a receptor binding region binds to a receptor of a target cell. The receptor of the target cell is other than the amphotropic cell receptor. The receptor binding region may be a receptor binding region of a human virus. A portion of the envelope gene may be deleted and the deleted portion is replaced with another receptor binding region or ligand. Such vector particles are targetable to a desired target cell or tissue and may be administered directly to the desired target cell or tissue as part of a gene therapy procedure, or administered directly into the patient.		

-1-

TARGETABLE VECTOR PARTICLES

This invention relates to "targetable" vector particles. More particularly, this invention relates to vector particles which include a receptor binding region that binds to a receptor of a target cell of a human or non-human animal.

Vector particles are useful agents for introducing gene(s) or DNA (RNA) into a cell, such as a eukaryotic cell. The gene is controlled by an appropriate promoter. Examples of vectors which may be employed to generate vector particles include prokaryotic vectors, such as bacterial vectors; eukaryotic vectors, including fungal vectors such as yeast vectors; and viral vectors such as DNA virus vectors, RNA virus vectors, and retroviral vectors. Retroviral vectors which have been employed for generating vector particles for introducing genes or DNA (RNA) into a cell include Moloney Murine Leukemia Virus, spleen necrosis virus, and vectors derived from retroviruses such as Rous Sarcoma Virus and Harvey Sarcoma Virus. The term "introducing" as used herein encompasses a variety of methods of transferring genes or DNA (RNA) into a cell, such methods including transformation, transduction, transfection, and infection.

Vector particles have been used for introducing DNA (RNA) into cells for gene therapy purposes. In general, such a procedure involves obtaining cells from a patient and using a vector particle to introduce desired DNA (RNA) into the cells and

The envelope of murine leukemia viruses includes a protein known as gp70. Such viruses can be made "targetable" to a specific type of cell if a portion of the gp70 protein is deleted and replaced with a receptor binding region or a ligand which binds to a receptor of a target cell. Thus, in a preferred embodiment, there is provided a retroviral vector wherein a portion, but not all, of the DNA (RNA) encoding gp70 protein has been deleted and replaced with DNA (RNA) encoding a receptor binding region or a ligand which binds to a receptor of a target cell.

In general, gp70 protein includes the following regions: (i) the secretory signal or "leader" sequence; (ii) the receptor binding domain; (iii) the hinge region; and (iv) the body portion. Preferably, at least a portion of the DNA (RNA) encoding the receptor binding domain of gp70 protein is deleted and replaced with DNA (RNA) encoding a receptor binding region or a ligand which binds to a receptor of a target cell. More preferably, DNA (RNA) encoding the entire receptor binding domain of gp70 protein is deleted and replaced with DNA (RNA) encoding a receptor binding region or a ligand which binds to a receptor of a target cell. In another embodiment, DNA (RNA) encoding the entire receptor binding domain of gp70 protein, plus all or a portion of the DNA (RNA) encoding the hinge region of gp70 protein is deleted and replaced with DNA (RNA) encoding a receptor binding region or a ligand of a target cell.

The gp70 protein may be derived from an ecotropic murine leukemia virus, a xenotropic murine leukemia virus, or an amphotropic murine leukemia virus. Ecotropic gp70 (or eco gp70) (SEQ ID NO:1) is a protein having 469 amino acids, and is encoded by (SEQ ID:2). Amino acid residues 1-33 constitute the leader sequence; amino acid residues 34-263 constitute the receptor binding domain; amino acid residues 264-312 constitute the hinge region; and amino acid residues 313-469 constitute the body portion. Preferably, DNA (RNA) encoding at least a portion of

as orosomucoid, and asialofetuin. AGP is a natural high-affinity ligand for ASG-R. The asialoglycoprotein receptor, or ASG-R, is expressed only by hepatocytes. The receptor is present at about 3×10^5 copies per cell, and such receptors have a high affinity for asialoglycoproteins such as AGP. Thus, the engineering of retroviral vector particles to contain asialoglycoprotein in place of the natural receptor binding domain of gp70 generates retroviral vector particles which bind to the asialoglycoprotein receptor of hepatocytes, which provides for an efficient means of transferring genes of interest to liver cells.

Cell lines which generate retroviral vector particles that are capable of targeting the hepatocyte's asialoglycoprotein receptor without the removal of the particle's terminal sialic acid groups by neuraminidase treatment, can be developed by selection with the cytotoxic lectin, wheat germ agglutinin (WGA). Cell lines which express the retroviral proteins gag and pol become retroviral vector packaging cell lines after they are transfected with the plasmids encoding chimeric envelope genes. These cell lines express the corresponding chimeric gp 70 glycoproteins. Upon exposure to successively higher concentrations of WGA, the outgrowth of cells which synthesize glycoproteins that lack terminal sialic acid groups, is favored (Stanley, et al., Somatic Cell Genetics, Vol. 3, pgs. 391-405 (1977)). This selection permits the isolation of cells which synthesize oligosaccharides terminating in galactosyl sugar groups. Such cells will allow the construction of packaging cell lines that are capable of generating retroviral vector particles which target the asialoglycoprotein receptor. It is also possible to select subpopulations of packaging cells which have other distinct glycotypes, such cells yielding viral vectors that potentially are capable of targeting cells other than hepatocytes. Macrophages, for example, express unique, high-mannose receptors. The PHA-resistant subpopulation will have N-linked oligosaccharides which terminate in high-mannose

mammalian genome. Another measure of rarity or scarcity of a restriction enzyme site in mammals is its representation in mammalian viruses, such as SV40. In general, an enzyme whose recognition sequence is absent in SV40 may be a candidate for being a "rare" mammalian cutter.

Examples of restriction enzyme sites having an average frequency of appearance in eukaryotic genes of less than once in 10,000 base pairs include, but are not limited to the NotI, SnaBI, Sall, XhoI, ClaI, SacI, EagI, and SmaI sites. Preferred cloning sites are selected from the group consisting of NotI, SnaBI, Sall, and XhoI.

Preferably, the multiple cloning site has a length no greater than about 70 base pairs, and preferably no greater than about 60 base pairs. In general, the multiple restriction enzyme site, or multiple cloning site is located between the 5' LTR and 3' LTR of the retroviral vector. The 5' end of the multiple cloning site is no greater than about 895 base pairs from the 3' end of the 5' LTR, preferably at least about 375 base pairs from the 3' end of the 5' LTR. The 3' end of the multiple cloning site is no greater than about 40 base pairs from the 5' end of the 3' LTR, and preferably at least 11 base pairs from the 5' end of the 3' LTR.

Such vectors may be engineered from existing retroviral vectors through genetic engineering techniques known in the art such that the retroviral vector includes at least four cloning sites wherein at least two of the cloning sites are selected from the group consisting of the NotI, SnaBI, Sall, and XhoI cloning sites. In a preferred embodiment, the retroviral vector includes each of the NotI, SnaBI, Sall, and XhoI cloning sites.

Such a retroviral vector may serve as part of a cloning system for the transfer of genes to such retroviral vector. Thus, there may be provided a cloning system for the manipulation of genes in a retroviral vector which includes a retroviral vector including a multiple cloning site of the type hereinabove

targetable, whereby the receptor binding region enables the vector particles to bind to a target cell. The retroviral vector particles thus may be directly administered to a desired target cell ex vivo, and such cells may then be administered to a patient as part of a gene therapy procedure.

Although the vector particles may be administered directly to a target cell, the vector particles may be engineered such that the vector particles are "injectable" as well as targetable, i.e., the vector particles are resistant to inactivation by human serum, and thus the targetable vector particles may be administered to a patient by intravenous injection, and travel directly to a desired target cell or tissue without being inactivated by human serum.

The envelope of retroviruses also includes a protein known as p15E, and Applicants have found that retroviruses are susceptible to inactivation by human serum as a result of the action of complement protein(s) present in serum on the p15E protein portion of the retrovirus. Applicants have further found that such retroviruses can be made resistant to inactivation by human serum by mutating such p15E protein.

In one embodiment, therefore, the retroviral vector is engineered such that a portion of the DNA (RNA) encoding p15E protein (shown in the accompanying sequence listing as SEQ ID NO:7), has been mutated to render the vector particle resistant to inactivation by human serum; i.e., at least one amino acid but not all of the amino acids of the p15E protein has been changed or mutated.

p15E protein is a viral protein having 196 amino acid residues. In viruses, sometimes all 196 amino acid residues are present, and in other viruses, amino acid residues 181 to 196 (known as the "r" peptide), are not present, and the resulting protein is the "mature" form of p15E known as p12E. Thus, viruses can contain both the p15E and p12E proteins. p15E protein is anchored in the viral membrane such that amino acid

In one embodiment, the mutation of DNA (RNA) encoding p15 protein may be effected by deleting a portion of the p15E gene and replacing the deleted portion of the p15E gene with fragment(s) or portion(s) of a gene encoding another viral protein. In one embodiment, one portion of DNA encoding the p15 protein is replaced with a fragment of the gene encoding the p21 protein, which is an HTLV-I transmembrane protein. HTLV-I virus has been found to be resistant to binding by complement proteins and thus HTLV-I is resistant to inactivation by human serum (Hoshino, et al., Nature, Vol. 310, pgs. 324-325 (1984)). Thus in one embodiment, there is also provided a retroviral vector particle wherein a portion of the p15E protein has been deleted and replaced with a portion of another viral protein, such as a portion of the p21 protein.

p21 protein (as shown in the accompanying sequence listing as SEQ ID NO:8) is a protein having 176 amino acid residues, and which, in relation to p15E, has significant amino acid sequence homology. In one embodiment, at least amino acid residues 39 to 61, and 101 to 123 are deleted from p15E protein, and replaced with amino acid residues 34 to 56 and 96 to 118 of p21 protein. In one alternative, at least amino acid residues 39 to 123 of p15E protein are deleted and replaced with amino acid residues 34 to 118 of p21 protein.

In another embodiment, amino acid residues 39 to 69 of p15E protein are deleted and replaced with amino acid residues 34 to 64 of p21 protein, and amino acid residues 96 to 123 of p15E protein are deleted and replaced with amino acid residues 91 to 118 of p21 protein.

Vector particles generated from such packaging lines, therefore, are "targetable" and "injectable," whereby such vector particles, upon administration to a patient, travel directly to desired target cell or tissue.

The targetable vector particles are useful for the introduction of desired heterologous genes into target cells ex

Simplex virus particles), or synthetic particles may be constructed such that the vector particles include a receptor binding region that binds to a receptor of a target cell, where the receptor of a human target cell is other than the amphotropic cell receptor. Such vector particles are suitable for in vivo administration to a desired target cell.

Advantages of the present invention include the ability to provide vector particles which may be administered directly to desired target cell or tissues, whereby desired genes are delivered to the target cell or tissue, whereby the target cell or tissue may produce the proteins expressed by such genes.

This invention will now be described with respect to the following examples; however, the scope of the present invention is not intended to be limited thereby.

Example 1

Plasmid pCee (Figure 1), which contains the ecotropic murine leukemia virus gp70 and p15E genes under the control of a CMV promoter, was cut with AccI, and an AccI fragment encoding amino acid residues 1-312 of the *eco* gp70 protein was removed. Cloned into the AccI site was a PCR fragment containing the *eco* gp70 secretion signal (or leader, which includes amino acid residues 1-33 of *eco* gp70), followed by mature rabbit alpha-1 acid glycoprotein (amino acid residues 19-201) (Ray, et al., Biochemical and Biophysical Research Communications, Vol. 178, No. 2, pgs. 507-513 (1991)). The amino acid sequence of rabbit alpha-1 acid glycoprotein is shown in (SEQ ID NO:5), and the DNA sequence encoding therefor is shown in (SEQ ID NO:6). The resulting plasmid pAGP-1 (Figure 2) contains the *eco* gp70 leader sequence (amino acid residues 1-33 of *eco* gp70), a sequence encoding the mature rabbit alpha-1 acid glycoprotein (amino acid residues 19-201), and a sequence encoding amino acid residues 3 to 469 of *eco* gp70.

Example 2

Example 3

Plasmid pUC18RSVXeno (Figure 4), which contains the xenotropic murine leukemia virus gp70 and p15E genes under the control of an RSV promoter, was cut with AccI and StuI, and an AccI-StuI fragment encoding amino acid residues 1-258 of xeno gp70 was removed. Cloned into this site was a PCR generated AccI-StuI fragment encoding the xeno gp70 leader (amino acid residues 1-30), and the mature rabbit alpha-1 acid glycoprotein. The resulting plasmid, pAX2 (Figure 5), thus contains a sequence encoding the xeno gp70 leader, a sequence encoding the mature rabbit alpha-1 acid glycoprotein, and amino acid residues 259-443 of xeno gp70.

Example 4

Plasmid pUC18RSVXeno was cut with AccI and ClaI, and a fragment encoding amino acid residues 1-210 of xeno gp70 was removed. Cloned into this site was a PCR generated AccI-ClaI fragment encoding the xeno gp70 leader, followed by mature rabbit alpha-1 acid glycoprotein. The resulting plasmid, pAX6 (Figure 6), thus includes a sequence encoding the xeno gp70 leader, a sequence encoding mature rabbit alpha-1 acid glycoprotein, and amino acid residues 211-443 of xeno gp70.

Example 5

5×10^5 GPL cells on 10 cm tissue culture plates were transfected (using CaPO_4) with 30 $\mu\text{g}/\text{plate}$ of one of plasmid pAGP-1, pAGP-3, pAX2, or pAX6. The CaPO_4 is removed 24 hours later and 10 ml of fresh D10 medium is added for another 24 hours. The D10 medium is then removed and replaced with serum free DX medium for another 24 hours. The DX medium is then collected, filtered, and stored on ice. This supernatant contains the vector particles.

The supernatants were then filtered and collected by standard procedures and then centrifuged. After centrifugation the virus pellets were reconstituted in a buffer containing 0.1M sodium acetate, 0.15M sodium chloride, and 2mM calcium chloride.

were plated, 1 ml of D10 was removed from the first well and 2 of neuraminidase-treated (or untreated as a control) viral supernatant containing Chimeric-1 or Chimeric-3 was added and mixed well. 200 ul from the first well was diluted into the 2 present in the second well, was then mixed; and then 200 ul from the second well was diluted into the 1.8 ml present in the third well, thereby giving approximate dilutions of 2/3, 1/15, and 1/150. 8 ug/ml of Polybrene was included in each well during transduction. The viral particles were left in contact with the cells overnight, followed by removal of media containing viral particles, and replaced with D10 containing 1,000 mg/ml of G418. The medium was changed with fresh D10 and G418 every 4 to 5 days as necessary. G418-resistant colonies were scored after 2 to 3 weeks.

Example 6

The pre-packaging cell line GP8, which expresses the retroviral proteins gag and pol, and the packaging cell lines derived from them which also express the chimeric gp70 glycoproteins encoded by the plasmids pAGP-1, pAGP-3, pAX2, or pAX6 were maintained in cell culture and exposed to successively higher concentrations of wheat germ agglutinin; starting with 15 ug/ml. The cell lines were maintained under WGA selection in cell culture for 6 to 8 weeks until populations resistant to 40-50 ug/ml WGA were obtained. The latter were then subjected to fluorescence-activated cell sorting using FITC-conjugated lectins to enrich for the cells expressing the desired mutant glycotype (e.g., FITC-Erythrina Cristagalli agglutinin for beta-D-galactosyl groups, and FITC-concanavalin A for alpha-D-mannosyl groups). Retroviral vector packaging and producer cell lines were then generated from the resulting populations by standard techniques.

It is to be understood, however, that the scope of the present invention is not to be limited to the specific embodiments described above. The invention may be practiced

PATAP697

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT: Anderson, W. French
Baltrucki, Leon F.
Mason, James M.

(ii) TITLE OF INVENTION: Targetable Vector Particles

(iii) NUMBER OF SEQUENCES: 8

(iv) CORRESPONDENCE ADDRESS:

(A) ADDRESSEE: Carella, Byrne, Bain, Gilfillan
Cecchi & Stewart
(B) STREET: 6 Becker Farm Road
(C) CITY: Roseland
(D) STATE: New Jersey
(E) COUNTRY: USA
(F) ZIP: 07068

(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: 3.5 inch diskette
(B) COMPUTER: IBM PS/2
(C) OPERATING SYSTEM: PC-DOS
(D) SOFTWARE: DW4.V2

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER:
(B) FILING DATE:
(C) CLASSIFICATION:

(vii) PRIOR APPLICATION DATA:

(2) INFORMATION FOR SEQ ID NO: 1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 469 bases

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(ix) FEATURE:

(A) NAME/KEY: Ecotropic gp70 Protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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Met Ala Arg Ser Thr Leu Ser Lys Pro Leu
      5                               10
Lys Asn Lys Val Asn  Pro Arg Gly Pro Leu
      15                               20
Ile Pro Leu Ile Leu  Leu Met Leu Arg Gly
      25                               30
Val Ser Thr Ala Ser  Pro Gly Ser Ser Pro
      35                               40
His Gly Val Tyr Asn  Ile Thr Trp Glu Val
      45                               50
Thr Asn Gly Asp Arg  Glu Thr Val Trp Ala
      55                               60
Thr Ser Gly Asn His  Pro Leu Trp Thr Trp
      65                               70
Trp Pro Asp Leu Thr  Pro Asp Leu Cys Met
      75                               80
Leu Ala His His Gly  Pro Ser Tyr Trp Gly
      85                               90
Leu Glu Tyr Gln Ser  Pro Phe Ser Ser Pro
      95                               100
Pro Gly Pro Pro Cys  Cys Ser Gly Gly Ser

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275	280
Ser Lys Pro Lys Pro	Val Lys Ser Pro Ser
285	290
Val Thr Lys Pro Pro	Ser Gly Thr Pro Leu
295	300
Ser Pro Thr Gln Leu	Pro Pro Ala Gly Thr
305	310
Glu Asn Arg Leu Leu	Asn Leu Val Asp Gly
315	320
Ala Tyr Gln Ala Leu	Asn Leu Thr Ser Pro
325	330
Asp Lys Thr Gln Glu	Cys Trp Leu Cys Leu
335	340
Val Ala Gly Pro Pro	Tyr Tyr Glu Gly Val
345	350
Ala Val Leu Gly Thr	Tyr Ser Asn His Thr
355	360
Ser Ala Pro Ala Asn	Cys Ser Val Ala Ser
365	370
Gln His Lys Leu Thr	Leu Ser Glu Val Thr
375	380
Gly Gln Gly Leu Cys	Ile Gly Ala Val Pro
385	390
Lys Thr His Gln Ala	Leu Cys Asn Thr Thr
395	400
Gln Thr Ser Ser Arg	Gly Ser Tyr Tyr Leu
405	410
Val Ala Pro Thr Gly	Thr Met Trp Ala Cys
415	420
Ser Thr Gly Leu Thr	Pro Cys Ile Ser Thr
425	430
Thr Ile Leu Asn Leu	Thr Thr Asp Tyr Cys
435	440
Val Leu Val Glu Leu	Trp Pro Arg Val Thr

(2) INFORMATION FOR SEQ ID NO: 2:

(I) SEQUENCE CHARACTERISTICS

(A) LENGTH: 1446 bases

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(II) MOLECULE TYPE: viral DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

GGCTGCCGAC CCCGGGGGTG GACCATCCTC TAGACTGACA TGGCGCGTTA AACGCTCTCA
AAACCCCTTA AAAATAAGGT TAACCCGCGA GGCCCCCTAA TCCCCTTAAT TCTTCTGATG
CTCAGAGGGG TCACTACTGC TTGCCCCGGC TCCAGTCCTC ATCAAGTCTA TAATATCACC
TGGGAGGTAA CCAATGGAGA TCGGGAGACG GTATGGGCAA CTTCCTGGCAA CCACCTCTG
TGGACCTGGT GGCCTGACCT TACCCAGAT TTATGTATGT TAGCCACCA TGGACCATCT
TATTGGGGGC TAGAATATCA ATCCCCTTTT TCTTCTCCCC CGGGGCCCCC TTGTGTCTCA
GGGGGCAGCA GCCCAGGCTG TTCCAGAGAC TCGGAAGAAC CTTTAACCTC CCTCACCCCT
CGGTGCAACA CTGCTGGAA CAGACTCAAG CTAGACCAGA CAACTCATAA ATCAATGAG
GGATTTTATG TTTGCCCCGG GCCCCACCGC CCCCAGAAAT CCAAGTCATG TGGGGGTCCA
GACTCCTTCT ACTGTCCCTA TTGGGGCTGT GAGACAACCG GTAGAGCTTA CTGGAAGCCC
TCCTCATCAT GGGATTTTAT CACAGTAAAC AACAACTCTA CCTCTGACCA GGCTGTCCAG
GTATGCAAAG ATAATAAGTG GTGCAACCCC TTAGTTATTC GGTTTACAGA CGCCGGGAGA
CGGGTTACTT CCTGGACCAC AGGACATTAC TGGGGCTTAC GTTTGTATGT CTCGGACAA
GATCCAGGGC TTACATTTGG GATCCGACTC AGATACCAA ATCTAGGACC CCGCGTCCCA
ATAGGGCCAA ACCCCGTTCT GGCAGACCAA CAGCCACTCT CCAAGCCCCA ACCTGTTAAG
TCGCCTTCAG TCACCAAACC ACCCAGTGGG ACTCCTCTCT CCCCTACCCA ACTTCCACCG
GCGGGAACGG AAAATAGGCT GCTAACTTA GTAGACGGAG CCTACCAAGC CCTCAACCTC
ACCACTCTG ACAAACCCA AGAGTGCTGG TTGTGTCTAG TAGCGGGACC CCCCTACTAC

(2) INFORMATION FOR SEQ ID NO: 3:

(I) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 443 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(II) MOLECULE TYPE: protein

(ix) FEATURE:

- (A) NAME/KEY: xenotropic gp70 protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Met Glu Gly Ser Ala	Phe Ser Lys Pro Leu
5	10
Lys Asp Lys Ile Asn	Pro Trp Gly Pro Leu
15	20
Ile Val Met Gly Ile	Leu Val Arg Ala Gly
25	30
Ala Ser Val Gln Arg	Asp Ser Pro His Gln
35	40
Ile Phe Asn Val Thr	Trp Arg Val Thr Asn
45	50
Leu Met Thr Gly Gln	Thr Ala Asn Ala Thr
55	60
Ser Leu Leu Gly Thr	Met Thr Asp Thr Phe
65	70

Phe Thr Asp Ala Gly	Arg Lys Ala Ser Trp
195	200
Asp Ala Pro Lys Val	Trp Gly Leu Arg Leu
205	210
Tyr Arg Ser Thr Gly	Ala Asp Pro Val Thr
215	220
Arg Phe Ser Leu Thr	Arg Gln Val Leu Asn
225	230
Val Gly Pro Arg Val	Pro Ile Gly Pro Asn
235	240
Pro Val Ile Thr Asp	Gln Leu Pro Pro Ser
245	250
Gln Pro Val Gln Ile	Met Leu Pro Arg Pro
255	260
Pro His Pro Pro Pro	Ser Gly Thr Val Ser
265	270
Met Val Pro Gly Ala	Pro Pro Pro Ser Gln
275	280
Gln Pro Gly Thr Gly	Asp Arg Leu Leu Asn
285	290
Leu Val Glu Gly Ala	Tyr Gln Ala Leu Asn
295	300
Leu Thr Ser Pro Asp	Lys Thr Gln Glu Cys
305	310
Trp Leu Cys Leu Val	Ser Gly Pro Pro Tyr
315	320

(2) INFORMATION FOR SEQ ID NO: 4:

(I) SEQUENCE CHARACTERISTICS

(A) LENGTH: 1356 bases

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(II) MOLECULE TYPE: viral DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

GCGACAATC CTCCAGCCGG GAACAGCATG GAAGGTTTCTCAG CGTTCTCAAA ACCCCTTAAA
GATAAGATTA ACCCGTGGGG CCCCTAATA GTTATGGGGA TCTTGGTGAG GGCAGGAGCT
TCGGTACAAC GTGACAGCCC TCACCAGATC TTCAATGTTA CTTGGAGAGT TACCAACCTA
ATGACAGGAC AAACAGCTAA CGCCACCTCC CTCCTGGGGA CGATGACAGA CACCTTCCCT
AAACTATATT TTGACCTGTG TGATTAGTA GGAGACTACT GGGATGACCC AGAACCCGAT
ATTGGGGATG GTTGCCGCAC TCCCGGGGGA AGAAGAAGGA CAAGACTGTA TGACTTCTAT
GTTTGGCCCG GTCATACTGT ACCAATAGGG TGTGGAGGGC CGGGAGAGGG CTACTGTGGC
AAATGGGGAT GTGAGACCAC TGGACAGGCA TACTGGAAGC CATCATCATC ATGGGACCTA
ATTTCCCTTA AGCGAGGAAA CACTCCTAAG GATCAGGGCC CCTGTTATGA TTCCTCGGTC
TCCAGTGGCG TCCAGGGTCC CACACCGGGG GGTGGATGCA ACCCCCTGGT CTTAGAATTC
ACTGACGGCG GTAGAAAGCC CAGCTGGGAT GCCCCCAAAG TTTGGGACT AAGACTCTAT
CGATCCACAG GGGCCGACCC GGTGACCCGG TTCTCTTTGA CCCGCCAGGT CCTCAATGTA
GGACCCCGCG TCCCATTTGG GCCTAATCCC GTGATCACTG ACCAGCTACC CCCATCCCAA
CCCGTGCGA TCATGCTCCC CAGGCCTCCT CATCCTCCTC CTTCAGGCAC GGTCTCTATG
GTACCTGGGG CTCCTCCGCC TTCTCAACAA CCTGGGACGG GAGACAGGCT GCTAAATCTG
GTAGAAGGAG CCTACCAAGC ACTCAACCTC ACCAGTCTG ACAAACCCA AGAGTCTCTG
TTGTGTCTGG TATCGGGACC CCCCTACTAC GAAGGGCTTG CCGTCTAGG TACCTACTCC

(2) INFORMATION FOR SEQ ID NO: 5:

(1) SEQUENCE CHARACTERISTICS

- (A) LENGTH: 201 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS:
- (D) TOPOLOGY: linear

(11) MOLECULE TYPE: protein

(1x) FEATURE

- (A) NAME/KEY: rabbit alpha-1-acid glycoprotein

(x) PUBLICATION INFORMATION

- (A) AUTHOR Ray, et al.
- (B) TITLE:
- (C) JOURNAL: Biochem. and Biophys. Res. Comm.
- (D) VOLUME: 178
- (E) ISSUE: No. 2
- (F) PAGES: 507-513
- (G) DATE: 1991

(x1) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

Met Ala Leu Pro Trp	Ala Leu Ala Val Leu
5	10
Ser Leu Leu Pro Leu	Leu His Ala Gln Asp
15	20
Pro Ala Cys Ala Asn	Phe Ser Thr Ser Pro
25	30

Glu Ala Leu Thr Cys	Leu Gly Met Asn Lys
165	170
Thr Glu Val Val Tyr	Thr Asp Trp Thr Lys
175	180
Asp Leu Cys Glu Pro	Leu Glu Lys Gln His
185	190
Glu Glu Glu Arg Lys	Lys Glu Lys Ala Glu
195	200

Ser

ACAAGACGGA AGTCGTCTAC ACTGACTGGA CAAAGGATCT GTCCGAGCCG CTGGAGAAGC
 AACACGAGGA GGAGAGGAAG AAGGAAAAGG CAGAGTCATA GGGCACAGCA CCGGCTCCCG
 GACTCGGGGC CCACCCCTG CACCTGCCCTT TTGTTTGT TTGTAAATCT CTGTTCTTTC
 CCATGGTTGC ATCAATAAAA CTGCTGGACC AGTAAAAA

(2) INFORMATION FOR SEQ ID NO: 7:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 196 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS:

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(ix) FEATURE:

(A) NAME/KEY: ecotropic p15E protein.

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

Glu Pro Val Ser Leu Thr Leu Ala Leu Leu	
	5 10
Leu Gly Gly Leu Thr Met Gly Gly Ile Ala	
	15 20
Ala Gly Ile Gly Thr Gly Thr Thr Ala Leu	
	25 30
Met Ala Thr Gln Gln Phe Gln Gln Leu Gln	
	35 40
Ala Ala Val Gln Asp Asp Leu Arg Glu Val	
	45 50

Val Leu Thr Gln Gln Tyr His Gln Leu Lys

185

190

Pro Ile Glu Tyr Glu Pro

195

INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 176 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS:

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(ix) FEATURE:

(A) NAME/KEY: HTLV-I p21 protein

(x) PUBLICATION INFORMATION:

(A) AUTHOR: Malik, et al.

(B) TITLE:

(C) JOURNAL: J. Gen. Virol.

(D) VOLUME: 69

(E) ISSUE:

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Lys Asn His Lys Asn Leu Leu Lys Ile Ala

55 60

Gln Tyr Ala Ala Gln Asn Arg Arg Gly Leu

65 70

Asp Leu Leu Phe Trp Glu Gln Gly Gly Leu

75 80

Cys Lys Ala Leu Gln Glu Gln Cys Cys Phe

85 90

Leu Asn Ile Thr Asn Ser His Val Ser Ile

95 100

Leu Gln Glu Arg Pro Pro Leu Glu Asn Arg

105 110

Val Leu Thr Gly Trp Gly Leu Asn Trp Asp

115 120

Leu Gly Leu Ser Gln Trp Ala Arg Glu Ala

125 130

Leu Gln Thr Gly Ile Thr Leu Val Ala Leu

135 140

Leu Leu Leu Val Ile Leu Ala Gly Pro Cys

145 150

Ile Leu Arg Gln Leu Arg His Leu Pro Ser

155 160

Arg Val Arg Tyr Pro His Tyr Ser Leu Ile

165 170

Asn Pro Glu Ser Ser Leu

175

9. The vector particle of Claim 8 wherein said protein which binds to an asialoglycoprotein receptor of hepatocytes is alpha-1 acid glycoprotein.

10. The vector particle of Claim 1 and further including at least one heterologous gene.

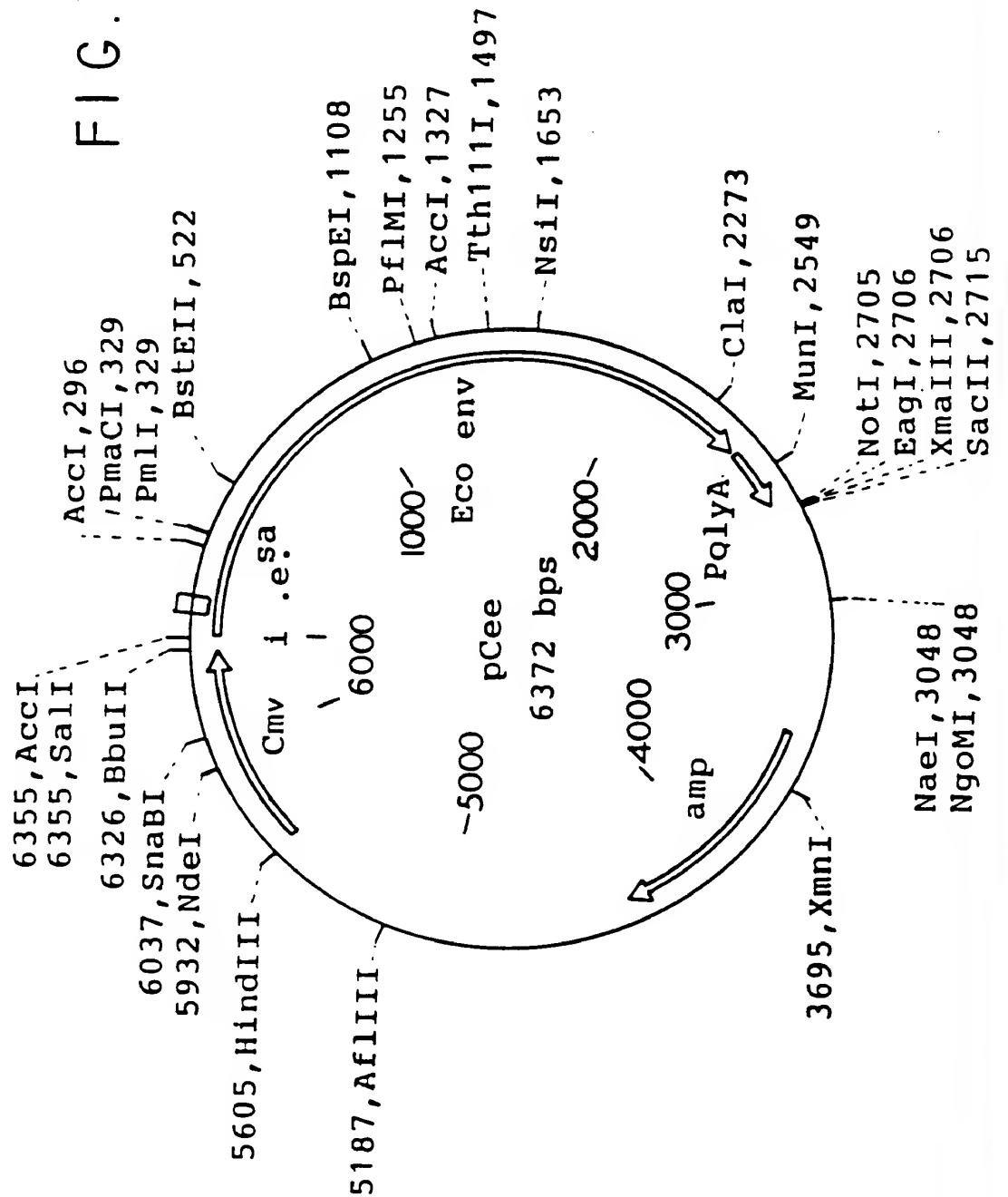
11. A method of introducing at least one heterologous gene into a target cell, comprising, administering to said target cell the vector particles of Claim 10.

12. The method of Claim 11 wherein said vector particles are administered ex vivo.

13. The method of Claim 11 wherein said vector particles are administered in vivo.

14. A packaging cell line which produces the retroviral particles of Claim 1.

FIG. 1



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FIG. 2

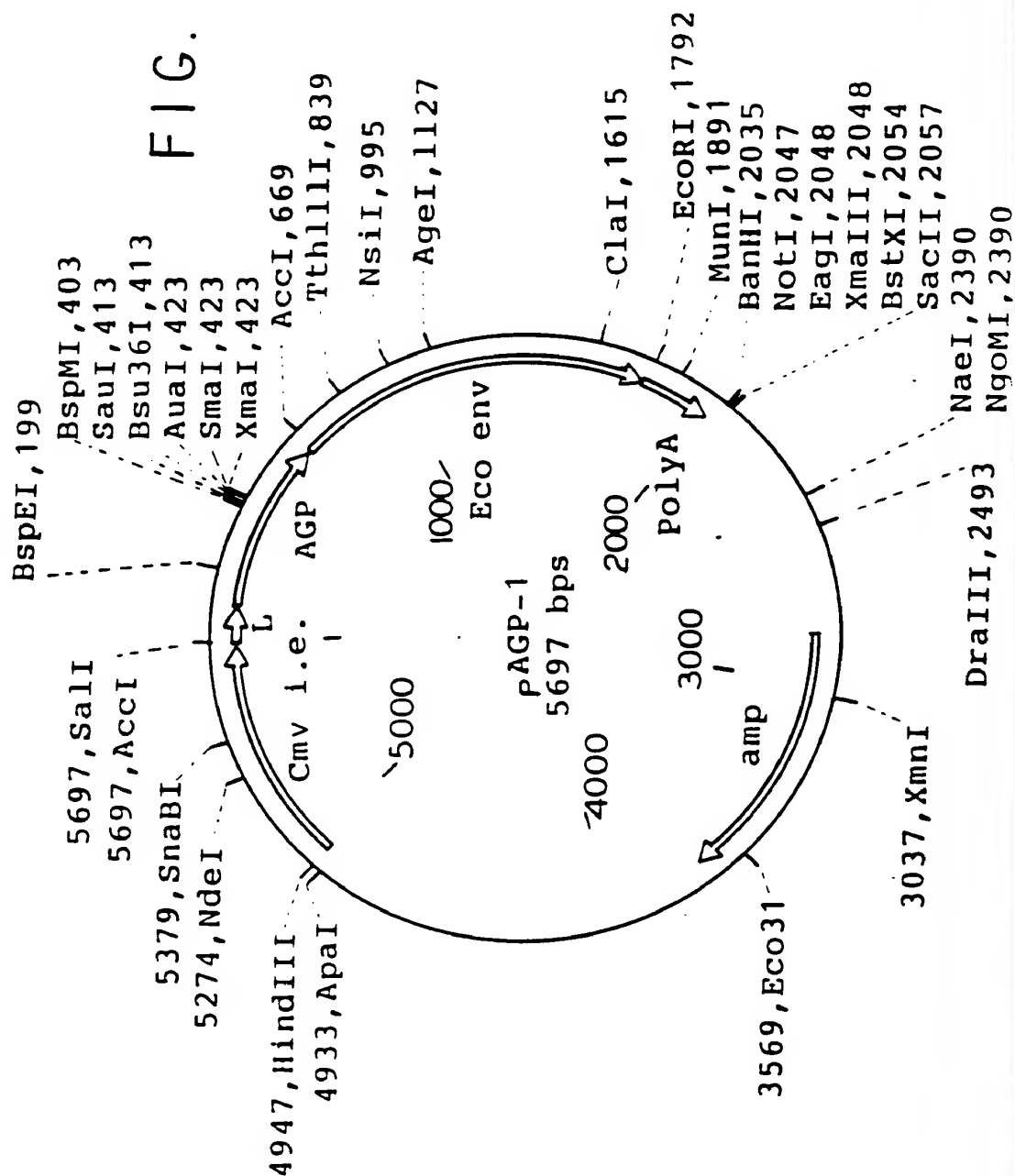




FIG. 4.

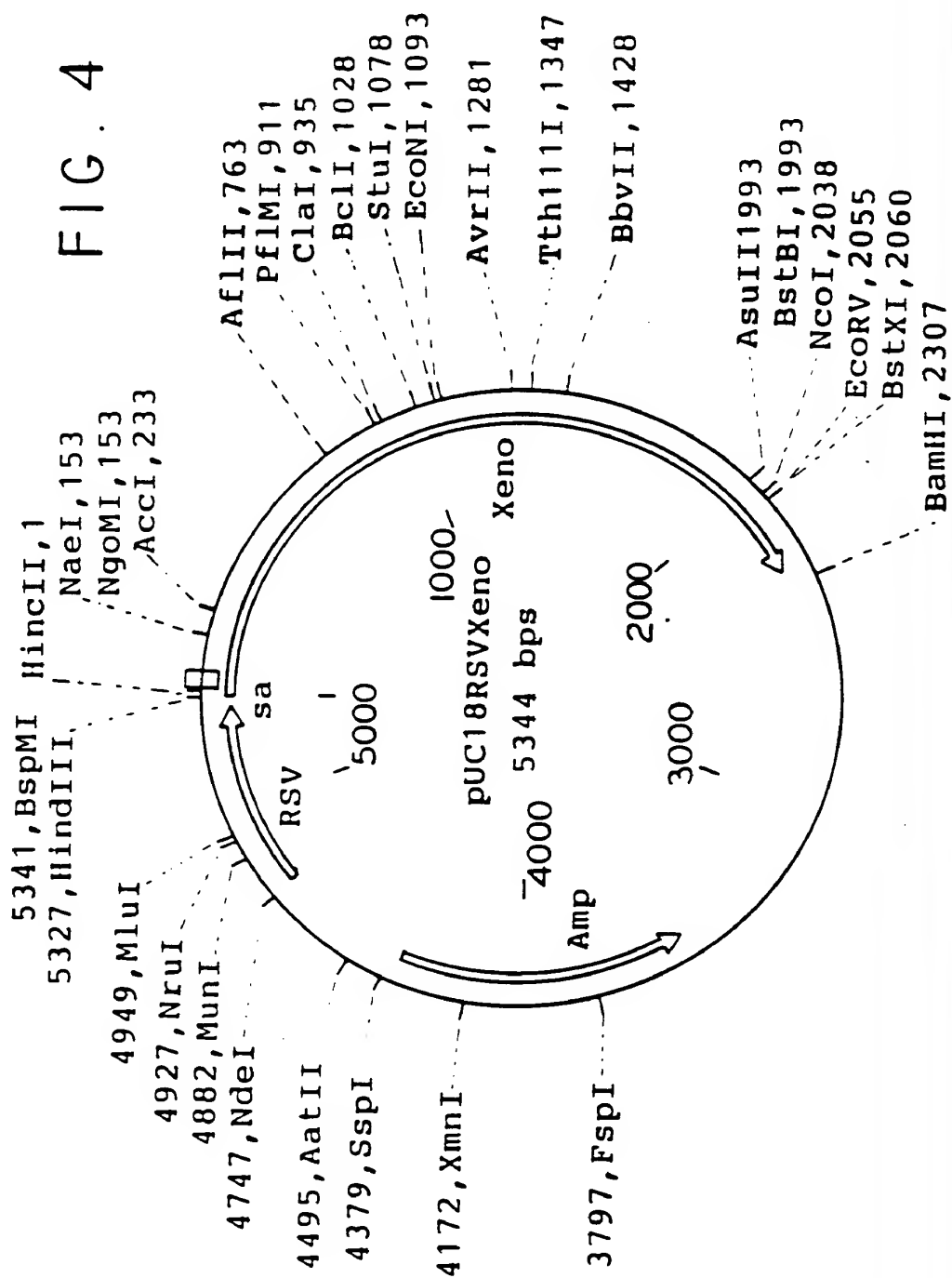
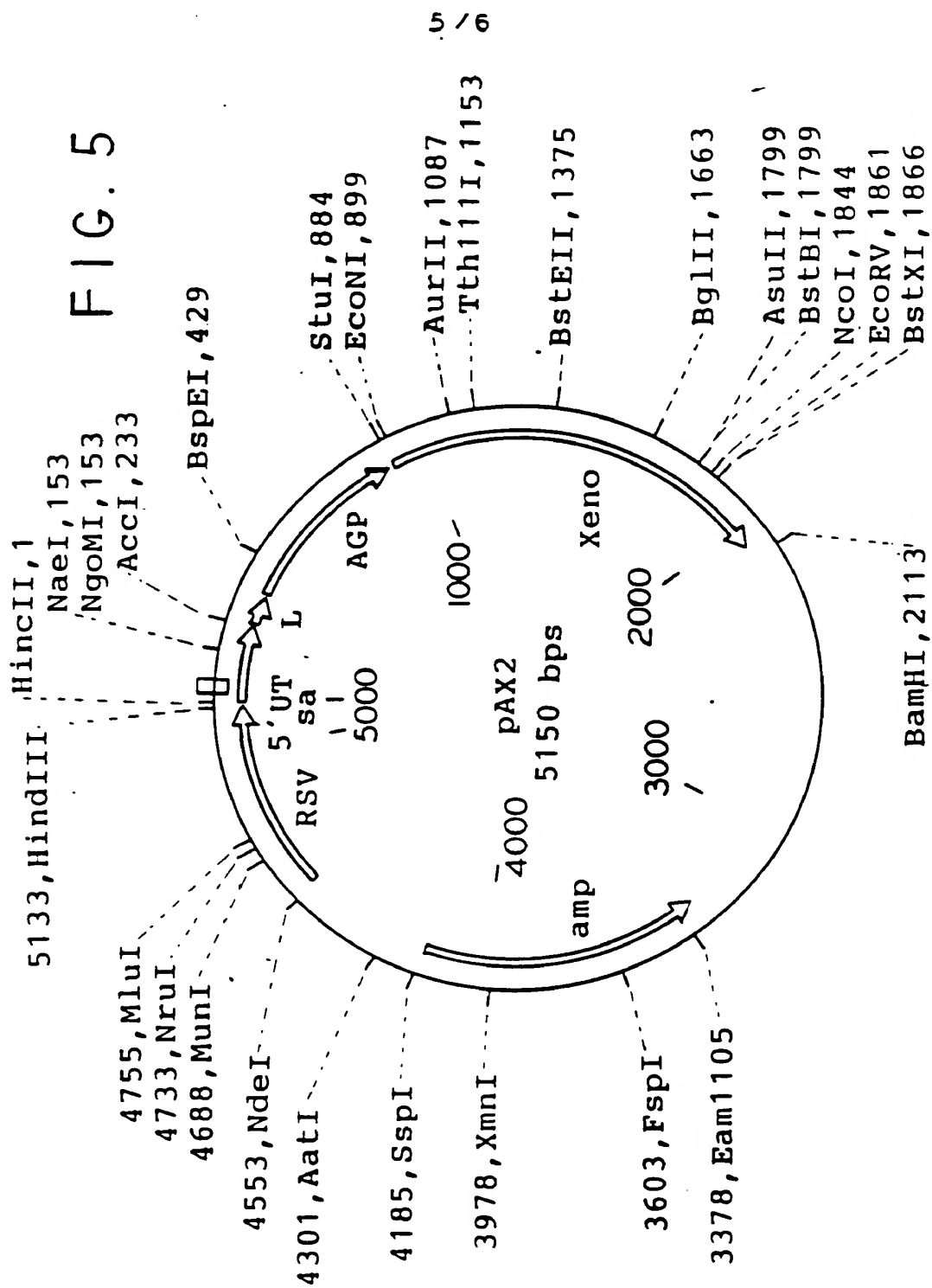
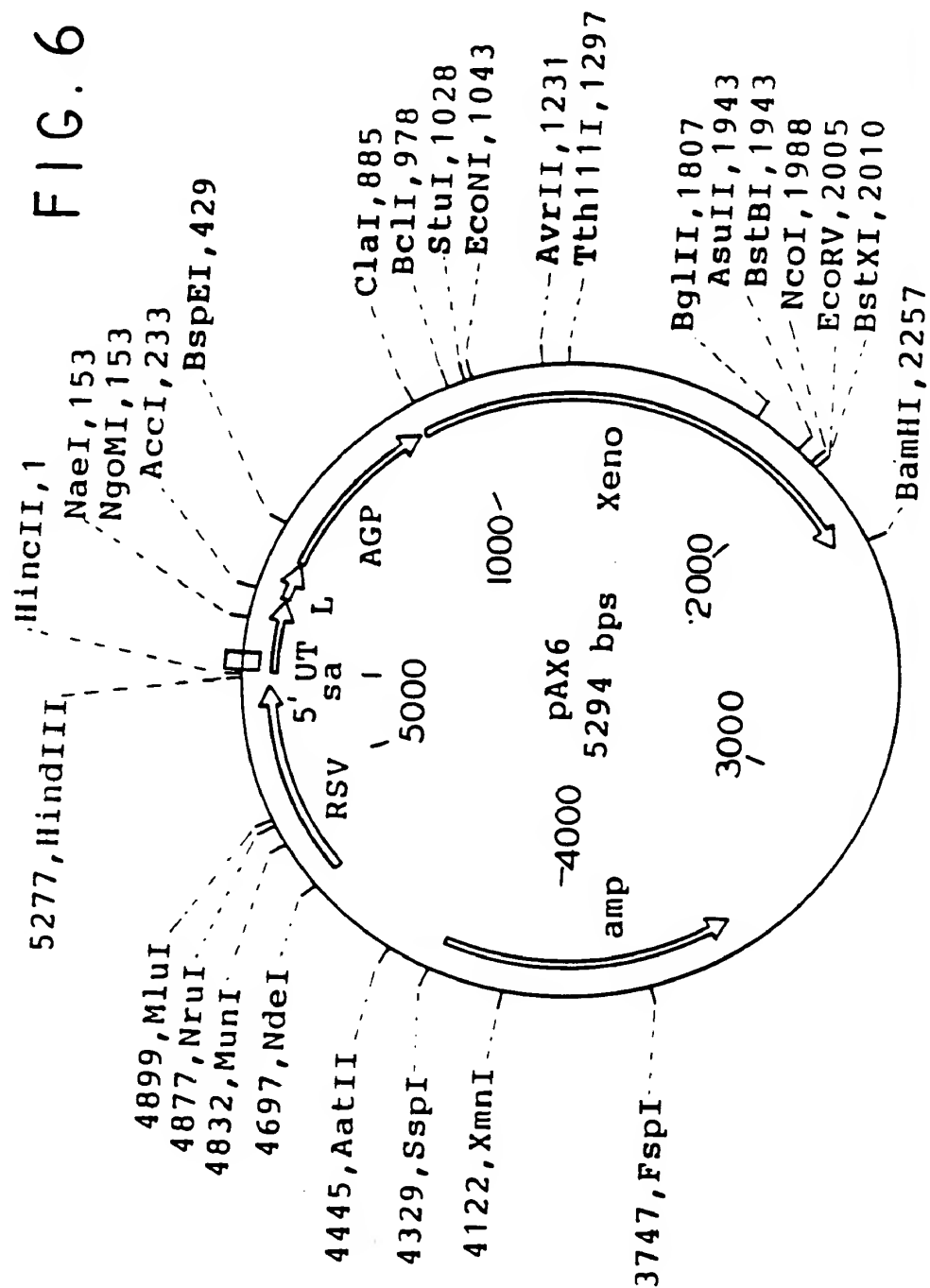


FIG. 5



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FIG. 6



INTERNATIONAL SEARCH REPORT

International application No.

PCT/US93/10522

A. CLASSIFICATION OF SUBJECT MATTER

IPC(5) : C12P 21/00; C12N 15/00, 15/58, 15/40, 15/48, 15/63, 15/86

US CL : 435/320.1, 69.1, 240.2; 424/93; 935/23, 32, 52, 57, 66, 70

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/320.1, 69.1, 240.2; 424/93; 935/23, 32, 52, 57, 66, 70

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS, Dialog, Biosis, Biotech, Medicine, Medline

Search Terms: retrovirus, vector, receptor, receptor binding protein

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	Journal of Virology, Volume 61, No. 5, issued May 1987, M.A. Bender et al., "Evidence that the Packaging Signal of Moloney Murine Leukemia Virus Extends Into the gag Region" pages 1639-1646, See particularly page 1640.	1-14
A	Biotechniques, Volume 7, No. 9, issued 1989, A.D. Miller et al., "Improved Retroviral Vectors for Gene Transfer and Expression" pages 980-990, See particularly page 984.	1-14

☐ Further documents are listed in the continuation of Box C.☐ See patent family annex.

Special categories of cited documents:	
* "A" documents defining the general state of the art which is not considered to be part of particular relevance	* "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
* "E" earlier document published on or after the international filing date	* "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
* "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another document or other special reason (as specified)	* "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
* "O" documents referring to an oral disclosure, use, exhibition or other means	* "A" document member of the same patent family
* "P" document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search

05 JANUARY 1994

Date of mailing of the international search report

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